Silver clusters shape determination from in-situ XANES data

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\textbf{ABSTRACT}

Knowledge of nanoparticle size, shape and morphology and of their in-situ transformations is crucial for establishing structure-properties relationship in nanosized materials that find applications, e.g., in plasmonic devices and heterogeneous catalysis. Here we demonstrate that this information can be extracted reliably from in-situ X-ray absorption near edge structure (XANES) data, by combining ab-initio XANES simulations and machine learning (artificial neural network (NN)) approaches. Here we use NN-XANES method to extract information about the size, shape and interatomic distances in silver clusters, and to monitor their changes during the temperature-controlled particle aggregation.

1. Introduction

In addition to composition, structure and size, the shape of metallic NPs can also affect their properties, such as their atomic dynamics and thermal properties (Roldan Cuenya et al., 2010, 2011), electronic state (Behafarid et al., 2012) and catalytic activity (Mostafa et al., 2010; Roldan Cuenya, 2012; Narayanan and El-Sayed, 2004; Grunwaldt et al., 2000), plasmonic and optical properties (Tao et al., 2008; Kelly et al., 2003). Ability to determine reliably NPs shape is thus a step towards establishing structure-properties relationship in these materials. Important in this regard are in-situ studies of NPs shape, since NPs structure, size and shape can change in their expected working conditions, e.g., due to NPs coalescence or Ostwald ripening (Porsgaard et al., 2012; Newton et al., 2007) or interactions with the support and adsorbates (Grunwaldt et al., 2000; Newton et al., 2007; Hansen et al., 2002). The set of tools that enable accurate in-situ determination of NPs shape is, however, limited. (Roldan Cuenya, 2012).

Among those few, X-ray absorption spectroscopy (XAS) stands out as a premier method that is sensitive to the details of atomic arrangements and that can be employed in a broad range of experimental conditions (Van Bokhoven and Lamberti, 2016). Sensitivity of XAS to contributions of distant shells and multiple scattering effects. This results in correlations in fitting variables and makes the procedure unstable. Such analysis of NPs shape is thus mostly limited to low-temperature studies of specially prepared model NPs, where EXAFS data of very good quality can be acquired.

Recently we demonstrated that for metallic particles CNs for the first few coordination shells can also be extracted from X-ray absorption near edge structure (XANES) by employing artificial neural network (NN) method (Timoshenko et al., 2017, 2018; Roese et al., 2018). Due to better signal-to-noise ratio in this portion of absorption spectra, good quality XANES data can be obtained in a broader range of experimental conditions, for more diluted samples and with better time resolution. XANES-based analysis is thus better suited for in-situ probing of NPs morphology. Here we apply NN method for interpretation of XANES data, collected in-situ during temperature-controlled aggregation of silver clusters embedded in room temperature ionic liquid (RTIL). Preliminary results for this system for the first coordination shell were published before (Roese et al., 2018). Here we focus on the analysis of distant coordination shells, and, for the first time, we employ our method for determination of NPs shape from their in-situ XANES spectra.

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1 ppm Ag clusters in RTIL was sample A and (Fig. 2) gives us con-

Theoretical simulations con-

structures as in Refs. Timoshenko et al. (2017), Timoshenko et al.

For Ag K-edge spectra in bulk Ag both codes provide good agreement

Ag NPs (Montano et al., 1984), atom coordinates in the structure

input vector is a discretized XANES spectrum. As the output we use CNs

WEff (Rehr et al., 2010) and FDMNES (Bunau and Joly, 2009) codes.

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NN is a composite function that can be trained to represent almost any

To account for possible shortening of interatomic distances in small

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The sample with \( \approx 1 \) ppm Ag clusters in RTIL was filled into a liquid sample holder with Kapton windows, and its temperature was controlled using a circulation thermostat. XAS data for separated clusters were collected at 250 K. For this temperature cluster aggregation is not observed on a timescale of one day (according to UV/vis data) (Roese et al., 2018). Afterwards the sample was warmed up to room temperature, and UV/vis data were collected for 2.5 h in total during the aggregation process. In addition to XAS data for the sample at the end of this aggregation process, we also collected XAS for an “intermediate state” after 80 min at room temperature. Each time the sample was cooled again to 250 K to avoid aggregation during the XAS measurement. More details on sample preparation and characterization are given in Roese et al. (2018).

In addition, we analyze also XAS data for two reference samples: Ag foil and sub-nanometer Ag cluster sample (\( \text{Ag}_{\text{DNA}} \)), prepared by DNA assisted synthesis (Gwinn et al., 2008; O’Neill et al., 2009). XAS data for the latter sample were collected at X18A beamline at NSLS facility. According to the indirect fluorescence measurements, \( \text{Ag}_{\text{DNA}} \) sample contains Ag clusters with 11–13 atoms.

2. Method description and validation

We follow the same procedure as in Ref. Timoshenko et al. (2017). NN is a composite function that can be trained to represent almost any relationship between NN input and output vectors by tuning the weights of different NN nodes (LeCun et al., 2015). In our case, the input vector is a discretized XANES spectrum. As the output we use CNs in the first three coordination shells (\( C_1, C_2, C_3 \)), as well as the effective interatomic distance \( \tilde{R} \). For NN training we use a set of XANES spectra, for which the true values of the structure parameters (\( C_1, C_2, C_3, \tilde{R} \)), are known. For this purpose we rely on ab-initio XANES modeling using FEFF (Rehr et al., 2010) and FDMNES (Bunau and Joly, 2009) codes.

For XANES simulations for Ag NPs with different sizes and shapes we use the same structure models with close-packed and icosahedral structures as in Refs. Timoshenko et al. (2017), Timoshenko et al. (2018). Theoretical simulations confirm the presence of particle size-effect in Ag K-edge XANES (Fig. 1), but it is relatively weak, especially for clusters larger than 1 nm. The non-linear sensitivity of NN to input features is thus crucial for reliable extraction of structural information.

To account for possible shortening of interatomic distances in small Ag NPs (Montano et al., 1984), atom coordinates in the structure models are rescaled to have nearest-neighbor distances in the range between 2.5 and 2.9 Å. In total, ca. 12,000 unique site-specific Ag K-edge XANES spectra were generated, which were further linearly fit in Ag K-edge XANES (Fig. 1), but it is relatively weak, especially for clusters larger than 1 nm. The non-linear sensitivity of NN to input features is thus crucial for reliable extraction of structural information.

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To validate the accuracy of the trained NN we apply it to another set of theoretical spectra for NPs of different sizes and shapes, which were not used for NN training. The good agreement between the known true

The validation of NN accuracy with simulated particle-averaged XANES for particles of different sizes and shapes and with experimental data for reference samples. For simulated spectra the obtained results are compared with the known true values of corresponding structure parameters. For experimental spectra NN-XANES results are compared with those from EXAFS fitting (Roese et al., 2018).

C1, C2, C3, R values for these models and NN yielded values \( \tilde{C}_1, \tilde{C}_2, \tilde{C}_3 \) and \( \tilde{R} \) (Fig. 2) gives us confidence that the information about these structure parameters indeed is present in XANES data and can be extracted by NN method.

3. Experimental data

Ag clusters with sizes 2.0 ± 0.6 nm were produced in a supersonic expansion process (Hövel et al., 1993) and deposited into ionic liquid (Roese et al., 2018; Engemann et al., 2016). Temperature-controlled aggregation of the clusters was monitored using UV/vis spectroscopy (Roese et al., 2018). To investigate the changes in cluster local structure and morphology, in-situ XAS measurements were performed.

XAS data (Fig. 3) were collected at beamline P64 at DESY facility. The sample with \( \approx 1 \) ppm Ag clusters in RTIL was filled into a liquid sample holder with Kapton windows, and its temperature was controlled using a circulation thermostat. XAS data for separated clusters were collected at 250 K. For this temperature cluster aggregation is not observed on a timescale of one day (according to UV/vis data) (Roese et al., 2018). Afterwards the sample was warmed up to room temperature, and UV/vis data were collected for 2.5 h in total during the aggregation process. In addition to XAS data for the sample at the end of this aggregation process, we also collected XAS for an “intermediate state” after 80 min at room temperature. Each time the sample was cooled again to 250 K to avoid aggregation during the XAS measurement.
A. C. is the statistical uncertainty of NN result). For sample, are significant.

Fig. 3. Experimental in situ XANES data for Ag clusters (separated, partially aggregated, aggregated). Experimental data for Ag foil and for Ag DNA sample are shown as references. Spectra are shifted vertically for clarity. In the insets - possible structure models for Ag NPs, constructed using CNs from NN-XANES analysis.

4. Results and discussion

To demonstrate the accuracy of NN-XANES method, we first apply it to reference spectra for Ag foil and Ag DNA sample. The obtained structure parameters yielded by NN are shown in Table 1. CNs and interatomic distances for bulk Ag are known from X-ray diffraction (Goon et al., 1957) and are also shown in Table 1. They agree well with the NN results. For Ag DNA sample, in turn, in addition to XANES data, also EXAFS data were collected that allowed us to obtain independently CN and interatomic distance for the first coordination shell (Roese et al., 2018). As shown in Fig. 2, EXAFS and NN-XANES results are in a good agreement. Slightly smaller R value for Ag DNA sample than for foil may be attributed to the shortening of interatomic distances with NPs size (Montano et al., 1984). CNs for the 2nd and 3rd coordination shells, yielded by NN for Ag DNA sample, are significantly smaller than those for bulk Ag, in agreement with the expected small NPs size. To check if C1, C2, and C3 values yielded by NN are realistic, we compare them with the CNs for a large set of NPs models that were used for NN training, and look for a model, for which $\sum_{i=1}^{N} (C_i - \bar{C}_i)^2 / \bar{C}_i^2$ is minimal (here $\bar{C}$ is the statistical uncertainty of NN result). For Ag DNA sample, the best agreement was obtained for cuboctahedral model with 13 atoms (Fig. 3). Corresponding CNs for this model are reported in Table 1, and for all coordination shells are indeed in a good agreement with NN result. The obtained model also agrees with the expected small size of Ag DNA NPs, as confirmed by fluorescence measurements. Consistency of the results observed for sub-nanometer NPs as well as for bulk Ag gives us confidence in the accuracy of our approach in a broad range of NPs sizes.

Finally, we discuss the results obtained from the in-situ XANES for separated and aggregated Ag clusters in RTIL. As shown in Fig. 3, these spectra are more similar to Ag foil spectrum, suggesting significantly larger NPs sizes in these samples than in Ag DNA sample. Signal-to-noise ratio is poor in these spectra, which prohibits advanced EXAFS-based analysis. CNs obtained by NN-XANES method and reported in Table 1, in turn, are reasonable. As expected, all CNs values are in-between those for Ag foil and Ag DNA samples. The obtained nearest-neighbor distance $R$ agrees with that for bulk Ag. The results, obtained for separated clusters, partially aggregated and completely aggregated NPs are very similar. Only a small increase in C1 value can be observed. C2 and C3 (which are more sensitive to NPs size and shape) (Frenkel et al., 2011; Jenitys, 1999) agree within error bars for all three cases. This is an indication that the NPs aggregation takes place without coalescence, and their local structure and shape are preserved during this process.

To identify a possible shape for Ag clusters in these samples, we perform similar analysis as was done above for Ag DNA sample. One possible shape for a “representative” NP model that has the same CNs as those yielded by the NN analysis (Table 1) is the truncated octahedral particle with 79 atoms (Fig. 3). The obtained NPs size (ca. 1.2 nm) is also close to the expected size for separated clusters (2.0 ± 0.6 nm) (Roese et al., 2018). One should emphasize here, however, that the experimental spectra are averaged over contributions of many NPs with, likely, quite different sizes and shapes. The shape analysis based on CNs extracted from XANES (or EXAFS) data thus can yield only an effective NP shape, and can be misleading, if the distribution of NPs sizes and shapes is broad (Frenkel et al., 2011).

5. Conclusions

We employed NN-XANES method for the analysis of NPs shape from in-situ XANES data, collected during NPs aggregation. Our results suggest that NPs aggregation takes place without coalescence and that the local structure and shape of these particles is preserved. This study shows that even when EXAFS data are not available due to experimental challenges, quantitative information on materials structure can be, nevertheless, extracted from XANES data, and can be used for advanced analysis, such as determination of NPs shape. We envision that our approach will be necessary in many cases, where the detailed knowledge of NPs structure and morphology is important, such as studies of heterogeneous catalysts and plasmonic materials.

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<table>
<thead>
<tr>
<th>Sample</th>
<th>C1</th>
<th>C2</th>
<th>C3</th>
<th>k (Å)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Foil</td>
<td>11.24(4)</td>
<td>5.7(4)</td>
<td>22.1(7)</td>
<td>2.883(7)</td>
</tr>
<tr>
<td>Ideal FCC</td>
<td>12</td>
<td>6</td>
<td>24</td>
<td>2.889 (Goon et al., 1957)</td>
</tr>
<tr>
<td>Ag DNA</td>
<td>5.3(2)</td>
<td>1.4(7)</td>
<td>8(2)</td>
<td>2.86(2)</td>
</tr>
<tr>
<td>Best model for Ag DNA</td>
<td>5.5</td>
<td>1.8</td>
<td>3.7</td>
<td></td>
</tr>
<tr>
<td>Separated clusters</td>
<td>8.3(3)</td>
<td>2.8(7)</td>
<td>14(1)</td>
<td>2.89(2)</td>
</tr>
<tr>
<td>Intermediate state</td>
<td>8.3(3)</td>
<td>1.4(7)</td>
<td>15(2)</td>
<td>2.91(1)</td>
</tr>
<tr>
<td>Aggregated clusters</td>
<td>8.6(2)</td>
<td>1.5(7)</td>
<td>14(1)</td>
<td>2.90(2)</td>
</tr>
<tr>
<td>Best model for separated/aggregated clusters</td>
<td>8.5(4)</td>
<td>3.2(3)</td>
<td>11.5</td>
<td></td>
</tr>
</tbody>
</table>

Table 1
Structure parameters obtained by NN from experimental XANES for reference samples (Ag foil and Ag DNA) and from in-situ XANES data for separated and aggregated Ag clusters in RTIL. Uncertainty of the last digit is given in parentheses. CNs corresponding to possible particle 3D models (Fig. 3) are also given for comparison.
References


