

# Growth and Maintenance of the 293FT Cell Line

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**User Manual** 

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## **Contents and Storage**

293FT Cell Line	The 293FT Cell Line is used for the production of lentiviral stocks. The 293FT Cell Line is supplied as one vial containing $3 \times 10^6$ frozen cells in 1 ml of Freezing Medium. <b>Upon receipt, store in liquid nitrogen until use.</b>		
CAUTION	Handle as potentially biohazardous material under at least Biosafety Level 2 containment. This product contains Dimethyl Sulfoxide (DMSO), a hazardous material. Review the Material Safety Data Sheet before handling.		

### **Accessory Products**

#### Accessory Products

The products listed below may be used with the 293FT Cell Line. For more information, refer to our Web site (www.invitrogen.com) or call Technical Support (see page 10).

Note: Some reagents are available in other sizes.

Item	Amount	Catalog no.
Lipofectamine <sup>™</sup> 2000 Reagent	0.75 ml	11668-027
	1.5 ml	11668-019
Dulbecco's Modified Eagle Medium	500 ml	11965-092
(D-MEM)	1000 ml	11965-084
Fetal Bovine Serum	100 ml	16000-036
	500 ml	16000-044
10 mM MEM Non-Essential Amino Acids Solution	100 ml	11140-050
200 mM L-Glutamine	100 ml	25030-081
MEM Sodium Pyruvate Solution (100X)	100 ml	11360-070
Penicillin-Streptomycin	100 ml	15070-063
Trypsin-EDTA	100 ml	25300-054
Geneticin <sup>®</sup> Selective Antibiotic	1 g	11811-023
	5 g	11811-031
	20 ml (50 mg/ml)	10131-035
	100 ml (50 mg/ml)	10131-027
Opti-MEM <sup>®</sup> I Reduced Serum Medium	100 ml	31985-062
	500 ml	31985-070
Phosphate-Buffered Saline (PBS), pH 7.4	500 ml	10010-023
	1 L	10010-031
Trypan Blue Stain	100 ml	15250-061

## Introduction

## **Product Information**

293FT Cell Line	The 293FT Cell Line is a very suitable host for lentiviral production. The 293FT Cell Line is derived from the 293F Cell Line (see below) and stably expresses the SV40 large T antigen from the pCMVSPORT6TAg.neo plasmid. Expression of the SV40 large T antigen is controlled by the human cytomegalo-virus (CMV) promoter and is high-level and constitutive. For more information about pCMVSPORT6TAg.neo, see the <b>Appendix</b> , page 9.
Use of the Cell Line	Studies have demonstrated maximal virus production in human 293 cells expressing SV40 large T antigen (Naldini <i>et al.</i> , 1996), making the 293FT Cell Line a particularly suitable host for generating lentiviral constructs using the ViraPower <sup>™</sup> Lentiviral Expression System available from Invitrogen (Catalog nos. K4950-00 and K4960-00).
Parental Cell Lines	The 293 Cell Line is a permanent line established from primary embryonal human kidney transformed with sheared human adenovirus type 5 DNA (Graham <i>et al.</i> , 1977; Harrison <i>et al.</i> , 1977). The E1A adenovirus gene is expressed in these cells and participates in transactivation of some viral promoters, allowing these cells to produce very high levels of protein.
	growing variant of the 293 cell line, and was originally obtained from Robert Horlick at Pharmacopeia.
Antibiotic Resistance	293FT cells stably express the neomycin resistance gene from pCMVSPORT6TAg.neo and should be maintained in medium containing Geneticin <sup>®</sup> at the concentration listed below. Expression of the neomycin resistance gene in 293FT cells is controlled by the SV40 enhancer/promoter.

## Methods

## **General Information**

General Cell Handling	Follow the general guidelines below to grow and maintain 293FT cells.				
	Make sure that all solutions and equipment that come in contact with the cells are sterile. Always use proper sterile technique and work in a laminar flow hood.				
	• Before starting experiments, be sure to have cells established and also have some frozen stocks on hand. We recommend using early-passage cells for your experiments.				
	<ul> <li>For general maintenance of cells, pass 293FT cells when they are &gt; 80% confluent. Avoid overgrowing cells before passaging.</li> </ul>				
	• Maintain 293FT cells in complete medium containing 500 μg/ml Geneticin <sup>®</sup> .				
	• Use trypan blue exclusion to determine cell viability. Log phase cultures should be >90% viable.				
	• When thawing or subculturing cells, transfer cells into medium warmed to room temperature.				
	• Cells should be at the appropriate confluence and at greater than 90% viability prior to transfection (see page 8).				
Before Starting	Be sure to have the following solutions and supplies available:				
	• 15 ml sterile, conical tubes				
	<ul> <li>Appropriate sized tissue culture flasks and pipettes</li> </ul>				
	Complete medium (see page 3)				
	• 50 mg/ml Geneticin <sup>®</sup>				
	Phosphate-Buffered Saline (PBS; Invitrogen, Catalog no. 10010-023)				
	Reagents for counting cells				
	Trypsin/versene (EDTA) solution or other trypsin solution				
	• Freezing Medium (see pages 3 and 7)				
	Table-top centrifuge				
	Cryovials (if needed)				

Continued on next page

## **General Information, continued**

Media for 293FT Cells	The table below lists the recommended complete medium, freezing medium, and antibiotic concentration required to maintain and culture the 293FT Cell Line. <b>Note:</b> Fetal bovine serum should not be heat-inactivated for use with the 293FT Cell Line.						
		Complete Medium	[Antibiotic]	]	Freezing Medium		
	D	-MEM (high glucose)	500 µg/ml	90% co	90% complete medium		
	10	)% fetal bovine serum (FBS)	Geneticin®	10% D	OMSO		
	0. A	mM MEM Non-Essential nino Acids (NEAA)					
	6	mM L-glutamine					
	1	l mM MEM Sodium Pyruvate					
	10	% Pen-Strep (optional)					
Preparing Medium	Th no glu Pro 0.1 glu	is will ensure that the concent t get too low over time due to te: 293FT cells grow well in 6 mM tamine may reduce growth. epare the complete D-MEM m . mM MEM Non-Essential Am atamine as described below us	ration of L-glutar its slow degradati I L-glutamine, but h edium containing nino Acids, 1 mM s sing reagents	nine in co ion. igher cor 10% FB sodium	omplete medium will neentrations of L- S supplemented with pyruvate and 2 mM L-		
	Perform all steps in a tissue culture hood under sterile conditions.						
	1.	1. Remove 100 ml D-MEM from 1 L D-MEM bottle and replace with 100 ml FBS.					
	2.	To the bottle of medium, add	d the following:				
		200 mM L-Glutamine (100X) 10 mM MEM Non-Essential 100 mM MEM Sodium Pyru <i>Optional:</i> Penicillin-Streptom	Amino Acids (100 vate (100X) avcin (100X)	)X)	10 ml 10 ml 10 ml 10 ml		
	3.	Filter sterilize the medium using 0.45 um filtration device.			vice.		
	4.	Store the complete medium at 4°C until use. The medium is stable for 6 months at 4°C (avoid introducing any contamination into the medium).			um is stable for 1 into the medium).		
	5.	To an aliquot of the complet medium with 500 μg/ml Ge	<mark>e medium, add Ge</mark> neticin <sup>®</sup> .	eneticin <sup>®</sup>	to prepare complete		

### **Thawing Cells**

Introduction	The 293FT Cell Line is supplied in a vial containing 3 x 10 <sup>6</sup> cells in 1 ml of Freezing Medium. Store frozen 293FT cells in liquid nitrogen until ready to use.			
CAUTION	Handle as potentially biohazardous material under at least Biosafety Level 2 containment. This product contains Dimethyl Sulfoxide (DMSO), a hazardous material. Review the Material Safety Data Sheet before handling.			
Thawing Cells	Use the following procedure to thaw 293FT cells to initiate cell culture. Thaw cells in prewarmed, complete medium <b>without</b> Geneticin <sup>®</sup> .			
	<ol> <li>Remove the vial of frozen cells from liquid nitrogen and thaw quickly in a 37°C water bath.</li> </ol>			
	2. Just before the cells are completely thawed, decontaminate the outside of the vial with 70% ethanol, and transfer the cells to a sterile 15 ml tube containing PBS. Briefly centrifuge the cells at 150-200 x g and resuspend the cells in 2 ml complete medium without Geneticin <sup>®</sup> .			
	3. Transfer the cells to T-75 cm <sup>2</sup> flask containing 10 ml of complete medium without Geneticin <sup>®</sup> .			
	<ol> <li>Incubate the flask overnight at 37°C for allowing the cells to attach to the bottom of the flask.</li> </ol>			
	5. The next day, aspirate off the medium and replace with fresh, complete medium containing 500 $\mu$ g/ml Geneticin <sup>®</sup> .			
	6. Incubate the cells and check them daily until the cells are 80-90% confluent.			
	7. Proceed to <b>Subculturing Cells</b> , next page.			
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We recommend subculturing cells for a minimum of 3 passages after thawing before use in other applications.

# Don't add Geneticin to cells that are coming from Cryogenic Storage

## Subculturing Cells

Introduction	Fo ce co	bllow the recommendation lls. Maintain cells as adher ntaining 500 μg/ml Genet	s and procedures in this section to subculture 293FT rent monolayer cultures in complete medium icin <sup>®</sup> .		
Subculturing Conditions		Use the following recommended conditions to subculture 293FT cells. For a procedure to subculture cells, see below.			
		Parameter	Recommended Condition		
	С	Cell density	$> 5 \times 10^5$ viable cells/ml (> 80% confluent)		
	С	Culture vessel	T-75 cm <sup>2</sup> to T-162 cm <sup>2</sup> disposable sterile T-flasks. Dilute cells in a total working volume of 15-20 ml for T-75 cm <sup>2</sup> flasks and 40-50 ml for T-162 cm <sup>2</sup> flasks		
	S	eeding density	2 to $5 \times 10^4$ viable cells/cm <sup>2</sup>		
	Ir	ncubation conditions	$37^{\circ}$ C incubator with a humidified atmosphere of $5-10\%$ CO <sub>2</sub> in air; loosen caps to allow for oxygenation/aeration		
Viability and Cell Density	try 1. 2. 3. 4. 5. 6. 7. [1. 8.	ypan blue exclusion metho Transfer a small aliquot dilute the cells such that than 100 or more than 1, To 1 ml of the diluted ce solution. Gently aspirate Record the dilution facto (amount of cell suspension amount of cell suspension Incubate the cells with the Count all cells (includin using a hemocytometer To calculate the total cell the dilution factor. To determine the viabili .00 - (Number of blue cells Cell viability should be	<ul> <li>of the cell suspension to a microcentrifuge tube and the total number of cells counted will not be less 000.</li> <li>ell suspension, add 100 µl Trypan Blue Stain (0.4%) e to mix.</li> <li>or. The dilution factor equals the total volume ion and amount of trypan blue) divided by the on.</li> <li>he trypan blue solution for 1-2 minutes.</li> <li>g the blue cells) using a Coulter Counter or manually chamber.</li> <li>ls per ml in suspension, multiply the total count by ty, count only the blue cells. Calculate the % viability:</li> <li>s ÷ Number of total cells)] x 100 at least 95% for healthy log-phase cultures.</li> </ul>		
		,	Continued on next page		
			10		

## Subculturing Cells, continued

Subculturing Cells	Use this procedure to subculture 293FT cells grown in a T-75 cm <sup>2</sup> flask. If you are using other-sized flasks, scale the reagent volumes accordingly.				
	1.	Remove all medium from the flask and wash the cells once with 10 ml PBS to remove excess medium and serum. Serum contains inhibitors of trypsin.			
	2.	Add 2 ml of trypsin/versene (EDTA) solution to the monolayer and incubate 1-5 minutes at room temperature until cells detach. Check the cells under a microscope and confirm that most of the cells have detached. If cells are still attached, incubate a little longer until most of the cells have detached.			
	3.	Add 8 ml complete medium containing Geneticin <sup>®</sup> and transfer the cell suspension to a 15 ml sterile, conical tube.			
	4.	Determine viable and total cell counts (see procedure above).			
	5.	Seed cells at the recommended density (see table on previous page), diluting in pre-warmed complete medium containing 500 $\mu$ g/ml Geneticin <sup>®</sup> . Incubate flasks as recommended (see table on previous page).			
	6.	Maintain cells as adherent monolayer cultures in complete medium containing 500 $\mu$ g/ml Geneticin <sup>®</sup> .			
	7.	For the transfection protocol, you will need $6 \ge 10^6$ 293FT cells for each sample (page 8).			

#### **Freezing Cells**

Introduction Once you have established the cells, we recommend freezing some cells for future use as described below. Freeze cells at a density of **at least** 3 x 10<sup>6</sup> viable cells/ml. Use a freezing medium composed of 90% complete medium and 10% DMSO. Prepare freezing medium immediately before use. Filter-sterilize the freezing medium and store at +4°C until use. Discard any remaining freezing medium after use. Freezing Cells Before starting, label cryovials and prepare freezing medium (see above). Keep the freezing medium on ice. 1. Culture the desired quantity of 293FT cells to 70-90% confluency. 2. Remove the cells from the tissue culture flask(s) following Steps 1-3, Subculturing Cells, page 6. 3. Determine viable and total cell counts (see procedure on page 5) and calculate the volume of freezing medium required to yield a final cell density of  $\geq 3 \times 10^6$  cells/ml. 4. Prepare the required volume of freezing medium (see above). 5. Centrifuge the cells suspension (from Step 2) at 250 x g for 5 minutes in a table top centrifuge at room temperature. Carefully aspirate off the medium and resuspend the cell pellet in the pre-determined volume of chilled freezing medium. Dispense aliquots of this suspension (frequently mixing to maintain a 6. homogeneous cell suspension) into cryovials according to manufacturer's specifications. 7. Freeze cells in an automated, controlled-rate freezing apparatus or using a manual method following standard procedures. For ideal cryopreservation, the freezing rate should be a decrease of 1°C per minute. 8. Transfer vials to liquid nitrogen storage. Note: You may check the viability and recovery of frozen cells 24 hours after storing cryovials in liquid nitrogen by following the procedure outlined in Thawing Cells, page 4.

## Don't add Geneticin to cells that are for Cryogenic Storage

## **Transfecting Cells**

Transfection Methods	The 293FT Cell Line is generally amenable to transfection using standard methods including calcium phosphate precipitation (Chen & Okayama, 1987; Wigler <i>et al.</i> , 1977), lipid-mediated transfection (Felgner <i>et al.</i> , 1989; Felgner & Ringold, 1989), and electroporation (Chu <i>et al.</i> , 1987; Shigekawa & Dower, 1988). We typically use cationic lipid-based transfection reagents to transfect 293FT cells. Lipofectamine <sup>™</sup> 2000 is recommended, but other transfection reagents are suitable. Lipofectamine <sup>™</sup> 2000 is available from Invitrogen (see page vi for ordering information).
Transient Transfection	The 293FT Cell Line may be transiently transfected with any plasmid. General guidelines are provided below.
	• Make sure that cells are healthy at the time of plating. Overgrowth of cells prior to passaging can compromise their transfection efficiency.
	• On the day before transfection, plate cells such that they will be at the appropriate confluence at the time of transfection (see manufacturer's recommendations for the transfection reagent you are using). <b>Example:</b> If you are using Lipofectamine <sup>™</sup> 2000 as a transfection reagent, plate cells such that they will be 90-95% confluent at the time of transfection.
	• Transfect your plasmid construct into the 293FT Cell Line using the method of choice (see above).
	<ul> <li>After transfection, add fresh growth medium containing 500 μg/ml Geneticin<sup>®</sup> and allow the cells to recover for 24-48 hours before proceeding to assay for expression of your gene of interest.</li> </ul>
Generating Stable Cell Lines	293FT cells can be used as hosts to generate a stable cell line expressing your gene of interest from most plasmids (see <b>Note</b> below). Remember that the introduced plasmid must contain a selection marker other than neomycin resistance. Stable cell lines can then be generated by transfection and dual selection with Geneticin <sup>®</sup> and the appropriate selection agent.
Note	Since 293FT cells stably express the SV40 large T antigen, we <b>do not</b> recommend generating stable cell lines with plasmids that contain the SV40 origin of replication.

#### Appendix

#### Map of pCMVSPORT6TAg.neo

#### Description

The pCMVSPORT6TAg.neo plasmid is derived from pCMVSPORT6, which has been modified to include the following features:

- The neomycin resistance gene for stable selection in mammalian cells (Southern & Berg, 1982). Expression of the neomycin resistance gene is controlled by the SV40 early enhancer/promoter from which the SV40 origin of replication has been removed.
- The gene encoding the SV40 large T antigen to facilitate optimal virus production (*e.g.* Invitrogen's ViraPower<sup>™</sup> Lentiviral Expression System) and to permit episomal replication of plasmids containing the SV40 early promoter and origin. Expression of the SV40 large T antigen is controlled by the human cytomegalovirus (CMV) promoter.



## **Technical Support**

Web Resources	Visit the Inv	itrogen website at <u>www.invitro</u>	g <u>en.com</u> for:	
	• Technical resources, including manuals, vector maps and sequences, application notes, SDSs, FAQs, formulations, citations, handbooks, etc.			
	Comple	te technical support contact info	ormation	
	• Access t	o the Invitrogen Online Catalog		
	Addition	nal product information and sp	ecial offers	
Contact Us	For more in internationa	formation or technical assistanc l offices are listed on our websi	e, call, write, fax, or email. Additional te ( <u>www.invitrogen.com</u> ).	
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	incidental, in warranty is so implied, incl	direct or consequential loss or dar ole and exclusive. No other warrar uding any warranty of merchantal	nage whatsoever. The above limited aty is made, whether expressed or bility or fitness for a particular purpose.	

## **Purchaser Notification**

Introduction	Use of the 293FT Cell Line is covered under the licenses detailed below.
Information for European Customers	The 293FT Cell Line is genetically modified and carries the pUC-derived plasmid, pCMVSPORT6TAg.neo. As a condition of sale, use of this product must be in accordance with all applicable local legislation and guidelines including EC Directive 90/219/EEC on the contained use of genetically modified organisms.
Limited Use Label License No. 5: Invitrogen Technology	The purchase of this product conveys to the buyer the non-transferable right to use the purchased amount of the product and components of the product in research conducted by the buyer (whether the buyer is an academic or for-profit entity). The buyer cannot sell or otherwise transfer (a) this product (b) its components or (c) materials made using this product or its components to a third party or otherwise use this product or its components or materials made using this product or its components for Commercial Purposes. The buyer may transfer information or materials made through the use of this product o a scientific collaborator, provided that such transfer is not for any Commercial Purpose, and that such collaborator agrees in writing (a) not to transfer such materials to any third party, and (b) to use such transferred materials and/or information solely for research and not for Commercial Purposes. Commercial Purposes means any activity by a party for consideration and may include, but is not limited to: (1) use of the product or its components, whether or not such product or its components are resold for use in research. For products that are subject to multiple limited use label licenses, the terms of the most restrictive limited use label license shall control. Life Technologies Corporation will not assert a claim against the buyer of infringement of patents owned or controlled by Life Technologies Corporation which cover this product or its components was employed, provided that neither this product nor any of its components was used in the manufacture of such product. If the purchaser is not willing to accept the limitations of this limited use statement, Life Technologies is willing to accept the limitations of this limited use statement, Life Technologies is willing to accept the limitations of the islinited use statement, Life Technologies is willing to accept the limitations of the similate use statement, Life Technologies is willing to accept the product or the technology embedded in it for any u

#### References

- Chen, C., and Okayama, H. (1987) High-Efficiency Transformation of Mammalian Cells by Plasmid DNA. Mol. Cell. Biol. 7, 2745-2752
- Chu, G., Hayakawa, H., and Berg, P. (1987) Electroporation for the Efficient Transfection of Mammalian Cells with DNA. Nucleic Acids Res. 15, 1311-1326
- Felgner, P. L., Holm, M., and Chan, H. (1989) Cationic Liposome Mediated Transfection. Proc. West. Pharmacol. Soc. 32, 115-121
- Felgner, P. L. a., and Ringold, G. M. (1989) Cationic Liposome-Mediated Transfection. Nature 337, 387-388
- Graham, F. L., Smiley, J., Russell, W. C., and Nairn, R. (1977) Characteristics of a Human Cell Line Transformed by DNA from Human Adenovirus Type 5. J. Gen. Virol. *36*, 59-74
- Harrison, T., Graham, F., and Williams, J. (1977) Host-range Mutants of Adenovirus Type 5 Defective for Growth in HeLa Cells. Virology 77, 319-329
- Naldini, L., Blomer, U., Gage, F. H., Trono, D., and Verma, I. M. (1996) Efficient Transfer, Integration, and Sustained Long-Term Expression of the Transgene in Adult Rat Brains Injected with a Lentiviral Vector. Proc. Natl. Acad. Sci. USA 93, 11382-11388
- Shigekawa, K., and Dower, W. J. (1988) Electroporation of Eukaryotes and Prokaryotes: A General Approach to the Introduction of Macromolecules into Cells. BioTechniques *6*, 742-751
- Southern, P. J., and Berg, P. (1982) Transformation of Mammalian Cells to Antibiotic Resistance with a Bacterial Gene Under Control of the SV40 Early Region Promoter. J. Molec. Appl. Gen. 1, 327-339
- Wigler, M., Silverstein, S., Lee, L.-S., Pellicer, A., Cheng, Y.-C., and Axel, R. (1977) Transfer of Purified Herpes Virus Thymidine Kinase Gene to Cultured Mouse Cells. Cell *11*, 223-232

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